

Evaluation of Temperate Maize Inbred Lines for Iron (Fe) Concentration using Atomic Absorption Spectrophotometry

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Maize (*Zea mays* L.) is one of the important cereal crops, ranking third after wheat and rice. It holds the title of the "Queen" of cereals due to its wide cultivation and nutritional value it provides. Especially iron is crucial for the proper functioning of haemoglobin and blood formation. Unfortunately, insufficient consumption of iron-rich foods is the primary cause of iron deficiency in the underdeveloped countries. A research study was conducted at the Department of Plant Breeding and Genetics, University of Agriculture, Faisalabad, aiming to investigate the diversity of 20 indigenous maize (*Zea mays* L.) inbred lines. Seeds were sown in a Randomized Complete Block Design (RCBD) with three replications. The study was conducted under open field conditions, focusing on genetic variations related to quality and yield traits. The results were analysed using analysis of variance (ANOVA) to determine variability among the inbred lines. Various parameters were recorded at maturity and after harvesting i.e., plant height, ear traits, and grain iron content. The evaluation of grain iron content revealed significant variation among the inbred lines tested. The inbred line with the highest iron content was 6162, recording 40.083 µg/g. Additionally, the analysis of other morphological and yield traits showed significant variations among the inbred lines. Three lines showed promising iron content: 6162 (40.083 µg/g), CRT-3 (36.417 µg/g), and 6199 (35.25 µg/g). Based on these research findings, there is a clear need to develop a breeding plan to enhance iron content in local maize germplasm.

Keywords: Maize, Iron deficiency, Field conditions, Diversity, Grain iron, Breeding plan.

INTRODUCTION

Maize (*Zea mays* L.) is third important cereal crop after wheat and rice (FAO, 2021), and often said to be as "Queen" of cereals. It is widely grown throughout the world for its edible grains (Duvick, 2005). It is member of Poaceae family and genus *Zea*, which also includes other important cereals like sorghum and teosinte (Hufford *et al.*, 2011). The diploid number of chromosomes of maize are $2n = 2x = 20$, having a large genome size of 2.5 million base pairs (Schnable *et al.*, 2009). Maize plants are characterized by their tall, grass-like appearance, with narrow and long leaves and prominent tassel at the top of plant (Chandler and Brendel, 2002). It is a monoecious plant, having separate male and female flowers on the same plant i.e., pollens grow in tassel and ovary develop in ear respectively while ear specialized structure for grain development (Lrsh and Nelson', 1989). Maize is an

outcrossing crop, depending on wind for pollination but sometimes insects also play a role in pollination (Stuber *et al.*, 1992). This is the reason why maize is genetically highly diverse crop, having a wide range of variation in characters such as yield, grain quality, and resistance to pests and diseases. Its grains can have a variety of colours, including yellow, white, red, and blue. The colour of grain is mainly determined by the presence of several pigments, such as carotenoids and anthocyanins. These pigmentation agents not only contribute towards the development of colour of grain, but also have health benefits for humans and animals that has maize grains as part of their food and feed (Harjes *et al.*, 2008). Maize is believed to be originated from Mesoamerica, which includes parts of Mexico and Central America. Domestication of maize crop dates to 8000 years by indigenous peoples (Matsuoka *et al.*, 2002). At present, maize is grown worldwide, with the United States, China and Brazil

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being the top three producers (FAO, 2020). It is the most important crop worldwide, in terms both economic value and its role as food for humans as well as animal feed. Among all the cereals there is an estimation of maize worldwide production about 1 billion metric tons per year. The genetic regulation of maize starch composition results in great diversity in the amylose to amylopectin ratio, making it ideal for a variety of industrial uses. Maize is also a great oil source, with a proven ability to lower blood cholesterol levels when taken by people. Maize has been used in a variety of ways across the world, resulting in the manufacture of several value-added products and fermented delicacies. Furthermore, maize naturally includes carotenoids such as beta-carotene, zeaxanthin, lutein, and cryptoxanthin, which have a variety of health advantages such as maintaining normal vision and reducing oxidative stress (Chaudhary *et al.*, 2014). Top corn producing countries include China, Brazil, United States of America, and Argentina (FAO, 2021). According to Pakistan Bureau of Statistics, Pakistan produces 6.6 million metric tons of corn from an area of approximately 1.1 million hectares. In Pakistan, maize is mostly cultivated in Punjab and Khyber Pakhtunkhwa provinces, Punjab is the major contributor to the country's total maize production. 0.5 % of gross domestic production of Pakistan is contributed by maize (GOP, 2022). For small scale farmers it is very important crop in Pakistan, specifically the areas where rain is the only sources of irrigation as it is the source of food feed and income. Nevertheless, maize production in Pakistan is facing a lot of challenges such as low yield, insufficient research and development, credit, and input access limitations as well as climate change-induced variability in rainfall patterns. Among all the mineral nutrients which are necessary in trace amounts for overall well-being of human beings as well as animals, Iron plays a paramount role in numerous metabolic events i.e., photosynthesis, respiration, chlorophyll synthesis, various redox reactions etc. (Briat and Lobréaux, 1997; Welch and Graham, 2004). Iron is a vital constituent of myoglobin and haemoglobin, playing their role in the supply and storage of oxygen. Most frequent dietary illness is anaemia, resulting from food that is iron deficient and troubling 32.9% of the world's population, with maximum risk of occurrence in southern Asia and sub-Saharan Africa (Wessells *et al.*, 2012; Kassebaum *et al.*, 2014). In females, during puberty as well as pregnancy, the highest iron deficiency effects (anaemia) can be seen. In the same way, iron deficiency disturbs cognitive growth of children until puberty. It also enhances the risk of getting infectious diseases as well as increased rate of mortality in children (Oliver and Gregory, 2015). Maize is a paramount constituent of various industrial products as well as animal feed (Nuss and Tanumihardjo, 2010), providing not only significant amounts of protein and energy, but also micronutrients for instance Zinc, Copper, and Iron (Shiferaw *et al.*, 2011; Dunn *et al.*, 2014). Iron (Fe) content in endosperm of grain of various

maize genotypes may fluctuate from 144 to 168 µg/g (Bityutskii *et al.*, 2001). Iron transporter genes are essential in this process since they oversee iron uptake, transport, and distribution within the plant (Curie and Briat, 2003). Iron transporter genes in maize have been found and characterized, including ZmYS1, ZmIRT1, ZmIRT2, and ZmFRO1 (Vigani *et al.*, 2013). These genes have been found to play critical roles in iron absorption and transport in several maize tissues (Schaaf *et al.*, 2004), but ZmIRT2 may strictly work together with ZmIRT1 uptake Fe in roots. In 21st century, worldwide great efforts have been made about the potential to produce food of greater nutritious value because cereal crops are genetically deprived of basic micronutrients (White and Broadley, 2009). Hence, there is an urgent requirement to cope this burning issue of micronutrient deficiencies that is attributable to what is referred to as "Hidden Hunger" and is affecting no less than 2 billion people, most of them living in Latin America, southern Asia as well as sub-Saharan Africa (McGuire, 2015). Fortification of food has an extensive account of use in developed world, it depends on the supplementation of micronutrients to the processed foods. However, process of food fortification is likely to have very prompt but less sustainable effects, because of several cost, safety, and technological concerns may prove to be limiting such intrusions (Allen *et al.*, 2006). Moreover, it is impossible to provide this kind of intervention to the target populations (Pfeiffer and McClafferty, 2007). "Hidden Hunger" due to micronutrient deficient diets, is a major public health issue prevailing all over developing world including Pakistan. Among all the micronutrient deficiencies iron deficiency is most prevalent leading to anaemia and other health issues. Iron deficiency in Bangladesh is a major health issue specifically in women and children. A study showed that the prevalence of anaemia due to iron deficiency remained 51.9 % overall- 47.4 % in urban and 53.1 % in rural regions respectively (Khan *et al.*, 2016). To deal with this public health issue worldwide, many approaches are being employed. These approaches include agronomic fortification, biofortification, breeding, and genetic engineering. Agronomic fortification involves addition of iron fertilizers to the soil to increase the iron content in crops (Bhardwaj *et al.*, 2022), this method has been used successfully in many countries, including Pakistan. Biofortification is another approach which includes breeding staple crops for higher iron contents. This approach has been successful in increasing the iron contents in many crops, including maize, rice, and beans (Cakmak, 2008). Connorton and Balk, (2019) studied that breeding for increased iron uptake efficiency has been extraordinarily successful in increasing iron contents of different crops so far. Genetic engineering is also being explored as a potential approach to increase the iron contents of crops. This involves modification of genes responsible for iron transport and storage in plants. Although this approach is in its early stages hence it has shown very promising results



in increasing the iron contents of rice and other crops (Lee *et al.*, 2009). Overall, a combination of all these approaches is likely to be the most effective way to address the iron deficiency and hidden hunger. These approaches can also have positive impacts on agricultural productivity and food security. By improving the amounts of micronutrients in the staple crops, micronutrient availability to the poor can be elevated, this will lead to the decrease in the predominance of micronutrient deficiencies. Plant breeding, as more sustainable approach, can be used to overcome the nutritional deficiencies, maize is a best food crop to reduce the prevalence of these nutritional deficiencies because it is one of the most affordable and easily available food crops. Hence, research was conducted under the agro-climatic conditions of Faisalabad.

MATERIALS AND METHODS

The current research was conducted at the Maize Research Area of Department of Plant Breeding and Genetics at University of Agriculture, Faisalabad. Coordinates of experimental area are 31°26'26" N latitude and 73°04'14" E longitude. Total twenty (20) maize inbred lines 6193, 6162, 6267, 6071, 6159, 6255, 6060, 6154, 2075, 6199, 349-A, 6266, 6276, 6212, 6180, 31(4B), CH-131265, PBG-1(2), CRT-3 and 6161, were obtained from the seed bank of the Department of Plant Breeding and Genetics. These genotypes were subjected to testing for evaluation of grain iron content and their morphological characteristics. Soil samples from experimental field were evaluated for iron content and the value of iron was 29.2 µg/g. The germplasm was planted in the mid of February 2022. The RH% in February, March, April, and May were 82.9%, 66.4%, 42.5%, and 43.4% respectively. The average temperatures for these months were 16.3, 25.2, 31.5, and 34.7 °C respectively. The experimental design used for this investigation was Randomised Complete Block Design (RCBD) with three (3) replications. P×P and R×R spaces were set at 20 cm and 75 cm, respectively. The inbred lines were assigned to the experimental units using a random number table. Total 12 to 14 irrigation cycles were done based on the moisture levels in the field and the needs of the crop. Fertilizer applications were made in accordance with crop demand. Plants were also self-pollinated when reached the appropriate stage.

preparation of samples for iron analysis:

Grinding: When the ears were ready, they were harvested from plants separately, then dried in a clean, shaded area to lower the moisture percentage of the harvested grains to 14%. To get representative samples of the grains, they were thoroughly ground into powder using a Cyclotech Sample Mill, which confirmed that there was no iron contamination.

Digestion: Ground material was digested using a 7:3 combinations of two acids (HNO₃: HClO₄). The first cycle

lasted sixty minutes at a temperature of 150 °C, followed by a thirty-five-minute cycle at a temperature of 235 °C.

Determination of iron (Fe) concentration through atomic absorption spectrophotometer: Minerals, especially iron (Fe), were measured in the prepared samples using an Atomic Absorption Spectrophotometer “Hitachi Polarised Zeeman AAS, Z-8200, Japan”, as defined in AOAC, (1990). The instrument’s operational parameters for these components are summarised in table (see Table 1).

Standards of preparation: Calibrated standards were generated by mixing an aqueous solution (1000 ppm) with a commercially accessible standard solution (Applichem®). The advancement of working standards necessitated the use of highly purified de-ionized water. All glass equipment used in the analysis were submerged in 8N HNO₃ for a long period of time before being cleaned with several changes of de-ionized water to ensure the best degree of precision.

Table 1. Conditions used to Determine Iron (Fe) by Atomic Absorption Spectrophotometer.

Factors	Values for Fe
Wavelength (nm)	248.3
Slit Width (nm)	0.2
Lamp Current (mA)	10.0
Burner Head	Standard type
Flame	Air-C ₂ H ₂
Burner Height (mm)	7.5
Oxidant gas pressure (Flow rate) (kpa)	160
Fuel gas pressure (Flow rate) (kpa)	6

RESULTS

Plant height: Analysis of variance expressed significant differences in plant height among the inbred lines (Table 2). Further investigation using mean values showed the range of plant heights which varied from 85.83 cm to 162.33 cm. Inbred line 6255 exhibited the highest plant height at 162.33 cm, while genotype CRT-3(6210) displayed the lowest 85.83 cm (Figure 1). Similar results were also stated by Nzuve *et al.* (2014).

Ear height: Analysis of variance expressed significant differences in ear height among the inbred lines (Table 2). Further investigation using mean values showed the range of ear heights which varied from 61.43 cm to 99.2 cm. Inbred line 6255 exhibited the highest plant height at 99.2 cm, while inbred line 6212 (CRT-3) displayed the lowest 61.43 cm (Figure 1). Similar results were also stated by Sherry *et al.* (2009).

Ear length: Analysis of variance expressed significant differences in ear length among the inbred lines (Table 2). Further investigation using mean values showed the range of ear length which varied from 6.03 cm to 15.3 cm. Inbred line 349 (A) exhibited the longest ear of 15.3 cm, while inbred line



Table 2. ANOVA of Various Traits.

Source	PH	EH	EL	ED	EW	GPR	HGWt	FeC
Rep (MS)	238.90	144.39	2.16	0.06	182.67	1.80	5.73	1.53
Inbred (MS)	1362.50	271.04	23.28	1.50	927.30	116.80	18.49	348.62
Error (MS)	91.78	60.22	1.00	0.31	138.17	9.92	7.05	2.33
F-value	14.85**	4.50**	23.18**	4.83**	6.71**	11.77**	2.62*	149.80**
P-value	0.000	0.000	0.000	0.000	0.000	0.000	0.006	0.000
CV%	7.67	10.46	9.04	12.07	10.27	11.62	13.45	7.71

6161 was recorded 6.03 cm long (Figure 1). Similar results were also stated by [Ojo et al. \(2007\)](#).

Ear diameter: Analysis of variance expressed significant differences in ear diameter among the inbred lines (Table 2). Further investigation using mean values showed the range of ear diameter which varied from 3.23 cm to 5.93 cm. Inbred line 6266 exhibited the highest ear diameter of 5.93 cm, while inbred line 6161 was recorded least diameter of 3.23 cm (Figure 1). Similar results were also stated by [Keskin et al. \(2005\)](#)

Ear weight: Analysis of variance expressed significant differences in ear weight among the inbred lines (Table 2). Further investigation using mean values showed the range of ear weight which varied from 71.33 g to 144.67 g. Inbred line 6180 exhibited the highest ear weight of 144.67 g, while inbred line 6159 recorded the least ear weight of 71.33 g (Figure 1). Similar results were also stated by [Sherry et al. \(2009\)](#).

Number of grains per row: Analysis of variance expressed significant differences in number of grains per row among the inbred lines (Table 2). Further investigation using mean values showed the range of number of grains per row which varied from 14.67 to 43.67 grains. Inbred line 6060 exhibited the highest number of grains per row 43.67, while inbred line 6161 recorded the least number of grains per row 14.67 (Figure 1). Similar results were also stated by [Ali et al. \(2014\)](#); [Zare et al. \(2011\)](#).

100 grain weight (g): Analysis of variance expressed significant differences in 100 grain weight among the inbred lines (Table 2). Further investigation using mean values showed the range of 100 grain weight which varied from 15.63 g to 22.87 g. Inbred line 6266 exhibited the highest 100 grain weight of 22.87 g, while inbred line 6255 recorded the least 100 grain weight of 15.63 g (Figure 1). Similar results were also stated by [Keskin et al. \(2005\)](#)

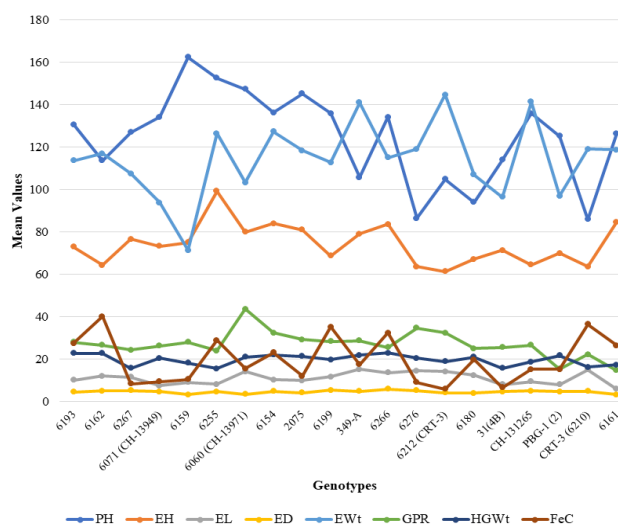
Grain iron (Fe) content ($\mu\text{g/g}$): Analysis of variance expressed significant differences in grain iron content among the inbred lines (Table 2). Further investigation using mean values showed the range of grain iron content which varied from 6 to 40.083 $\mu\text{g/g}$. Inbred lines with the highest grain iron contents were 6162 40.083 $\mu\text{g/g}$, followed by CRT-3 36.417 $\mu\text{g/g}$, and the inbred line with the lowest grain iron content was 6212 6.00 $\mu\text{g/g}$. According to a recent study by [Prasanna et al. \(2011\)](#), a genotype HP2 (Harvest Plus) showed 42.21

$\mu\text{g/g}$. Similar results were also stated by [Chakraborti et al. \(2011\)](#), [Phuke et al. \(2017\)](#), [Sharma et al. \(2021\)](#), [Qureshi et al. \(2021\)](#), [Saletnik et al. \(2022\)](#) and [Udo et al. \(2023\)](#).

Principle Component Analysis:

PC 1: PCA 1 accounts for a significant part of variability among inbred lines and traits which 31.3%. Traits like grains per row (GPR), hundred grain weight (HGWt), ear height (EH) and ear length (EL) contribute more to the variability captured by PCA 1. Inbred 6190 shows that it has trait values that are in positive association to ear length (EL) and hundred grain weight (HGWt). Inbred lines PBG1(2), 6255, 6161, 6212, 6276, and 349-A are apart from one another showing that they are variable from one another and have distinct trait profiles.

PC 2: PCA 2 accounts for relatively moderate amount of variability among inbred line and all the traits which is 18.8%. PCA 2 is influenced by plant height (PH), ear weight (EWt), ear diameter (ED) and grain iron content (FeC). There is slight degree of association between ear diameter (ED) and ear weight (EWt) as they are positioned closer. Inbred Lines 6193 and 6154 are extremely close showing that they share similar trait profiles. Inbred lines CH-131265, 6199, 6266, 6162, and CRT-3 are more scattered and apart from one another indicating that they have different trait profiles.

**Figure 1. Mean values of various traits.**

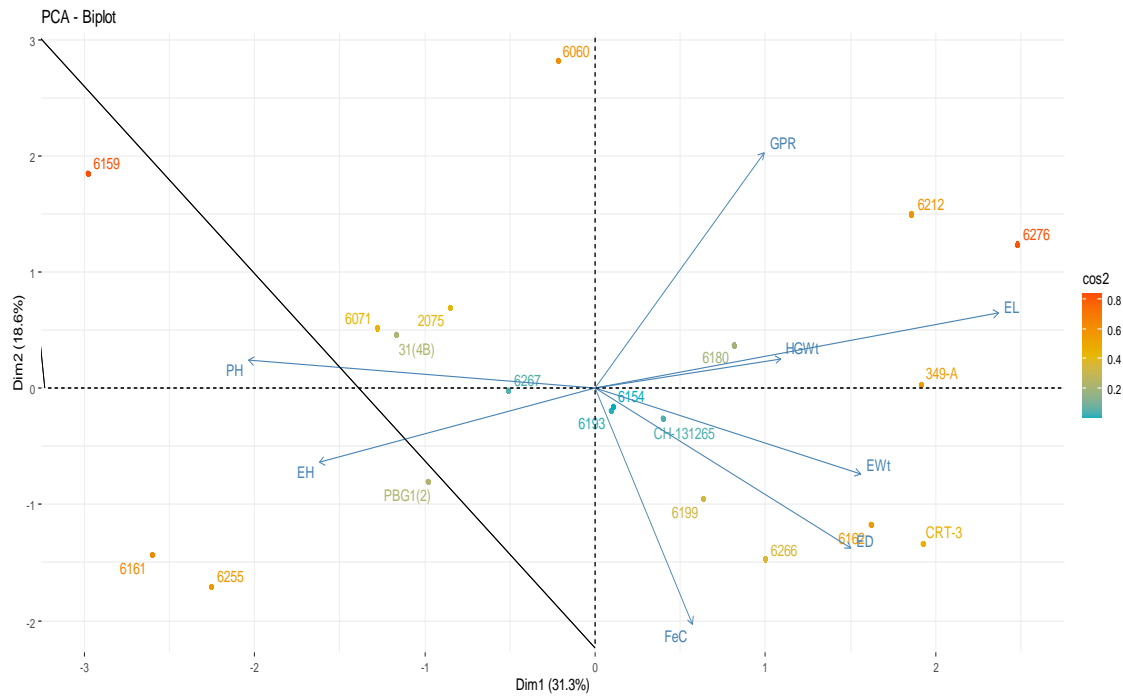


Figure 2. Principle Component Analysis - Biplot

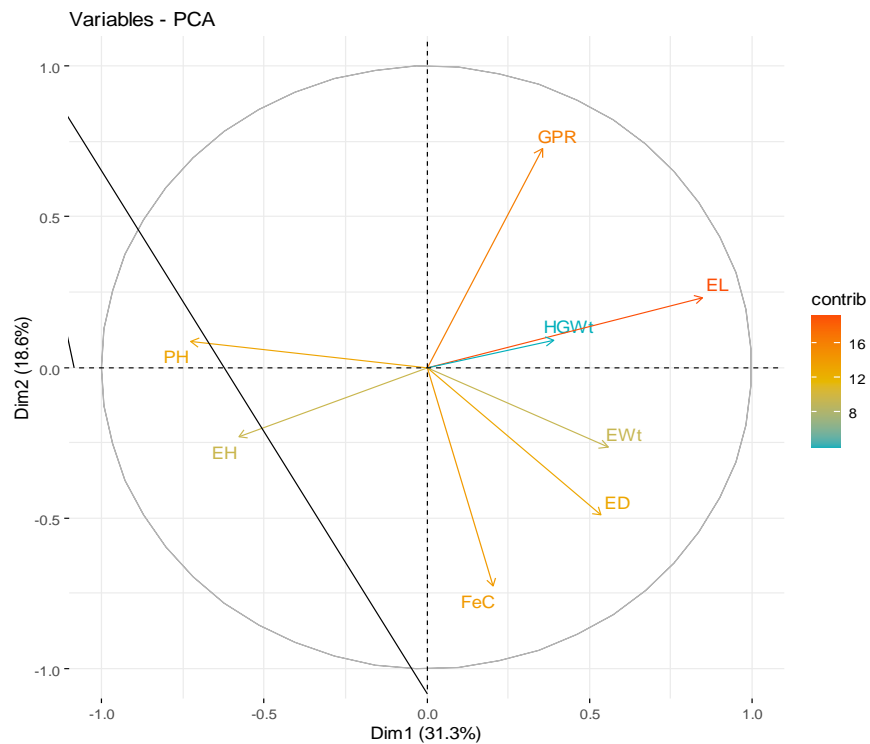


Figure 3. Variables – Principal Component Analysis

Conclusion: The outcomes of our investigation reveal notable iron concentrations in three distinct lines: 6162 (40.083 $\mu\text{g/g}$),

CRT-3 (36.417 $\mu\text{g/g}$), and 6199 (35.25 $\mu\text{g/g}$). These findings underscore the potential for targeted breeding initiatives



aimed at augmenting the iron content within indigenous maize germplasm. The imperative to formulate a comprehensive breeding strategy is evident, reflecting the significance of advancing nutritional traits in maize for broader agricultural and public health objectives.

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REFERENCES

Ali, Q., M. Ahsan, M. Hammad, N. Tahir, S. Maqsood and A. Basra. 2014. Gene action and Correlation Studies for Various Grain and its Contributing Traits in Maize (*Zea mays* L). Pakistan Journal of Botany 51:615-623

Allen, L., B. de Benoist, O. Dary and R. Hurrell. 2006. World Health Organization Report Part Guidelines on food fortification with micronutrients.

AOAC. 1990. Official methods of analysis. Association of official analytical chemists. Arlington, VA, USA.

Bhardwaj, A.K., S. Chejara, K. Malik, R. Kumar, A. Kumar and R.K. Yadav. 2022. Agronomic biofortification of food crops: An emerging opportunity for global food and nutritional security. Agronomy 14:220-226.

Bityutskii, N., S. Magnitski, I. Lapshina, E. Lukina, A. Soloviova and V. Patsevitch. 2001. Distribution of micronutrients in maize grain and their mobilisation during germination. Plant Nutrition 12:218-219.

Briat, J.F. and S. Lobréaux. 1997. Iron transport and storage in plants. Trends Plant Sciences 2:187-193.

Cakmak, I. 2008. Enrichment of cereal grains with zinc: Agronomic or genetic biofortification Plant Soil 302:1-17.

Chakraborti, M., B.M. Prasanna, F. Hossain, S. Mazumdar, A.M. Singh, S. Guleria and H.S. Gupta. 2011. Identification of kernel iron- and zinc-rich maize inbreds

and analysis of genetic diversity using microsatellite markers. Journal of Plant Biochemistry and Biotechnology 20:224-233.

Chandler, V.L. and V. Brendel. 2002. The maize genome sequencing project.

Chaudhary, D.P., S. Kumar and O.P. Yadav. 2014. Nutritive value of maize: Improvements, applications and constraints. p. 3-17. Maize: Nutrition dynamics and novel uses. Springer, India.

Connorton, J.M. and J. Balk. 2019. Iron biofortification of staple crops: Lessons and challenges in plant genetics. Plant Cell Physiology 60:1447-1456.

Curie, C. and J.F. Briat. 2003. Iron transport and signaling in plants. Annu. Rev. Plant Biology 54:183-206.

Dunn, M.L., V. Jain and B.P. Klein. 2014. Stability of key micronutrients added to fortified maize flours and corn meal. Ann. N. Y. Soil Science 1312:15-25.

Duvick, D.N. 2005. The contribution of breeding to yield advances in maize (*Zea mays* L.). p. 83-145. Advances in Agronomy. Academic Press.

FAO, 2020. Food and Agriculture Organization of the United Nations, Rome, Italy.

FAO, 2021. Food and Agriculture Organization of the United Nations, Rome, Italy.

Government of Pakistan (GOP), 2022. Economic Survey of Pakistan 2021-22. Islamabad.

Harjes, C.E., T.R. Rocheford, L. Bai, T.P. Brutnell, C.B. Kandianis, S.G. Sowinski, A.E. Stapleton, R. Vallabhaneni, M. Williams, E.T. Wurtzel, J. Yan and E.S. Buckler. 2008. Natural genetic variation in lycopene epsilon cyclase tapped for maize biofortification. Pakistan Journal of Botany 319:330-333.

Hufford, M.B., P. Gepts and J. Ross-Ibarra. 2011. Influence of cryptic population structure on observed mating patterns in the wild progenitor of maize (*Zea mays* spp. *parviglumis*). Molecular Ecology 20:46-55.

Kassebaum, N.J., R. Jasrasaria, M. Naghavi, S.K. Wulf, N. Johns, R. Lozano, M. Regan, D. Weatherall, D.P. Chou, T.P. Eisele, S.R. Flaxman, R.L. Pullan, S.J. Brooker and C.J.L. Murray. 2014. A systematic analysis of global anaemia burden from 1990 to 2010. Journal of Agronomy 123:615-624.

Keskin, B., I. Yilmaz and Ö. Arvas. 2005. Determination of Some Yield Characters of Grain Corn in Eastern Anatolia Region of Turkey. Journal of Agronomy 4:20-27.

Khan, J.R., N. Awan and F. Misu. 2016. Determinants of anaemia among 6-59 months aged children in Bangladesh: Evidence from nationally representative data. BMC 16:225-229.

Lee, S., U. Sil Jeon, S. Jin Lee, Y.-K. Kim, D. Pergament Persson, S. Husted, J.K. Schjørring, Y. Kakei, H. Masuda, N.K. Nishizawa and G. An. 2009. Iron fortification of rice seeds through activation of the



- nicotianamine synthase gene. *Proceed. National Academic Sciences* 106:22014-22019.
- Lrlish, E.E. and T. Nelson. 1989. Sex Determination in Monoecious and Dioecious Plants.
- Matsuoka, Y., Y. Vigouroux, M.M. Goodman, J. Sanchez G., E. Buckler and J. Doebley. 2002. A single domestication for maize shown by multilocus microsatellite genotyping. *Proceed. National Academic Sciences* 99:6080-6084.
- McGuire, S. 2015. FAO, IFAD, and WFP. The State of Food Insecurity in the World 2015: Meeting the 2015 International Hunger Targets: Taking Stock of Uneven Progress. Rome: FAO, 2015. *Advancements in Nutrition* 6:623-624.
- Nuss, E.T. and S.A. Tanumihardjo. 2010. Maize: A paramount staple crop in the context of global nutrition. *Compr. Rev. Food Science and Food Safety* 9:417-436.
- Nzuve, F., S. Githiri, D.M. Mukunya and J. Gethi. 2014. Genetic variability and correlation studies of grain yield and related agronomic traits in maize. *Journal Agriculture Sciences* 6:32-35.
- Ojo, G., D.K. Adedzwa and L.L. Bello. 2007. Combining ability estimates and heterosis for grain yield and yield components in maize (*Zea mays* L.). *J. Sustain. Develop. Agriculture Environment* 3:49-57.
- Oliver, M.A. and P.J. Gregory. 2015. Soil, food security and human health: a review. *Eur. J. Soil Science* 66:257-276.
- Pakistan Bureau of Statistics (PBS), 2022. Government of Pakistan, Islamabad, Pakistan.
- Pfeiffer, W.H. and B. McClafferty. 2007. HarvestPlus: Breeding crops for better nutrition. *Crop Science* 47: 88-105.
- Phuke, R.M., K. Anuradha, K. Radhika, F. Jabeen, G. Anuradha, T. Ramesh, K. Hariprasanna, S.P. Mehtre, S.P. Deshpande, G. Anil, R.R. Das, A. Rathore, T. Hash, B.V.S. Reddy and A. Ashok Kumar. 2017. Genetic variability, genotype \times environment interaction, correlation, and GGE biplot analysis for grain iron and zinc concentration and other agronomic traits in RIL population of Sorghum (*Sorghum bicolor* L. Moench). *Front. Plant Sciences* 8:712.
- Prasanna, B.M., S. Mazumdar, M. Chakraborti, F. Hossain, K.M. Manjaiah, P.K. Agrawal, S.K. Guleria and H.S. Gupta. 2011. Genetic variability and genotype \times environment interactions for kernel iron and zinc concentrations in maize (*Zea mays*) genotypes. *Indian Journal of Agriculture Sciences* 81:704-711.
- Qureshi, M.T., N. Iqbal, I.R. Noorka and H. Waheed. 2021. Bio-fortification of iron and zinc improves the biomass, uptake, distribution and yield of different maize varieties. *Journal of Plant Nutrition* 44:120-129.
- Saletnik, B., A. Saletnik, E. Słysz, G. Zagula, M. Bajcar, A. Puchalska-Sarna and C. Puchalski. 2022. The static magnetic field regulates the structure, biochemical activity, and gene expression of plants. *Molecules* 27:5823.
- Schaaf, G., U. Ludewig, B.E. Erenoglu, S. Mori, T. Kitahara and N. von Wirén. 2004. ZmYS1 functions as a proton-coupled symporter for phyto siderophore- and nicotianamine-chelated metals. *Journal of Biology and Chemistry* 279:9091-9096.
- Schnable, P.S., D. Ware, R.S. Fulton, J.C. Stein, F. Wei, S. Pasternak, C. Liang, J. Zhang, L. Fulton, T.A. Graves, P. Minx, A.D. Reily, L. Courtney, S.S. Kruchowski, W.R. McCombie, R.A. Wing and R.K. Wilson. 2009. The B73 maize genome: Complexity, diversity, and dynamics. *Sciences* 326:1112-1115.
- Sharma, D., R. Chhabra, V. Muthusamy, R.U. Zunjare and F. Hossain. 2021. Molecular characterization of elite maize (*Zea mays* L.) inbreds using markers associated with iron and zinc transporter genes. *Genet. Resour. Crop Evolution* 68:1545-1556.
- Sherry A, F.-G., E.S. Buckler, P. Tiffin, E. Ersoz and N.M. Springer. 2009. Heterosis Is Prevalent for Multiple Traits in Diverse Maize Germplasm. *PLoS One* 4:e7433-.
- Shiferaw, B., B.M. Prasanna, J. Hellin and M. Bänziger. 2011. Crops that feed the world 6. Past successes and future challenges to the role played by maize in global food security. *Food Security* 3:307.
- Šimić, D., R. Sudar, T. Ledenčan, A. Jambrović, Z. Zdunić, I. Brkić and V. Kovačević. 2009. Genetic variation of bioavailable iron and zinc in grain of a maize population. *Journal of Cereal Sciences* 50:392-397.
- Stuber, C.W., T. Stephen, E. Lincoln, D.W. Wolff, T. Helentjaris and E.S. Lander. 1992. Identification of genetic factors contributing to heterosis in a hybrid from two elite maize inbred lines using molecular markers. *Genetics* 132:823-839.
- Udo, E., A. Abe, S. Meseka, W. Mengesha and A. Menkir. 2023. Genetic analysis of zinc, iron and provitamin a content in tropical maize (*Zea mays* L.). *Agronomy* 13:266.
- Vigani, G., G. Zocchi, K. Bashir, K. Philippar and J.F. Briat. 2013. Cellular iron homeostasis and metabolism in plant. *Frontier in Plant Sciences* 4.
- Welch, R.M. and R.D. Graham. 2004. Breeding for micronutrients in staple food crops from a human nutrition perspective. *Journal of Experimental Botany* 55:353-364.
- Wessells, K.R., G.M. Singh and K.H. Brown. 2012. Estimating the global prevalence of inadequate zinc intake from national food balance sheets: effects of methodological assumptions. *PLoS One* 7:50565-.
- White, P.J. and M.R. Broadley. 2009. Biofortification of crops with seven mineral elements often lacking in human diets-iron, zinc, copper, calcium, magnesium, selenium and iodine. *New Phytology* 182:49-84.



- Xue, Y., S. Yue, W. Zhang, D. Liu, Z. Cui, X. Chen, Y. Ye and C. Zou. 2014. Zinc, iron, manganese and copper uptake requirement in response to nitrogen supply and the increased grain yield of summer maize. PLoS One. 9:93895-.
- Zare, M., R. Choukan, I. Heravan, M. Bihamta (Ghannadha) and K. Ordookhani. 2011. Gene action of some agronomic traits in corn (*Zea mays* L.) using diallel cross analysis. African Journal of Agriculture Research 6:20-26.

